A new species of *Euzetia* (Monogenea: Monocotylidae) on the gills of *Rhinoptera bonasus* (Rhinopteridae) from Ciudad del Carmen, Campeche, México

Una especie nueva de *Euzetia* (Monogenea: Monocotylidae) de las branquias de *Rhinoptera bonasus* (Rhinopteridae) de Ciudad del Carmen, Campeche, México

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Abstract. *Euzetia lamothei* n. sp. (Monogenea: Euzetiinae) is described from the gills of the stingray, *Rhinoptera bonasus*, collected from marine waters near Ciudad del Carmen, Campeche, Mexico. The species is assigned to *Euzetia* Chisholm and Whittington, 2001 because it has a haptor with 10 peripheral loculi and 1 central loculus flanked by 2 lateral loculi. The species described herein can be distinguished from the only known member of the genus, *E. occultum*, by the morphology of the male copulatory organ, the vas deferens sigmoid, and the presence of a vaginal chamber. *Euzetia lamothei* n. sp. was found on 1 of 7 *R. bonasus* (14%). This species is the first record of a member of the genus from the neotropics, the first from Mexico, and the second report of a monocotylid from *Rhinoptera*.

Key words: *Euzetia lamothei* n. sp., *Rhinoptera bonasus*, Euzetiinae, description, Campeche, Mexico.

Introduction

The subfamily Euzetiinae Chisholm and Whittington, 2001 was erected for *Euzetia occultum* Chisholm and Whittington, 2001, an exclusive parasite of *Rhinoptera neglecta* Ogilby, 1912 collected from Moreton Bay, Queensland, Australia (Chisholm and Whittington, 2001). During 2000 to 2004, as part of an ongoing study of helminth parasites of stingrays (Marques et al., 1995; Monks et al., 1996; Pulido-Flores and Monks, 2005) specimens of *R. bonasus* (Mitchill, 1815) were collected in coastal Mexican waters off the coasts of the states of Veracruz, Campeche, Yucatán, and Quintana Roo, Mexico. Of those collected near Ciudad del Carmen, Campeche, a single stingray harbored monogeneans on its gills with characteristics that indicated they should be assigned to *Euzetia* Chisholm and Whittington, 2001. These specimens are described herein.

Materials and methods

Seven specimens of *Rhinoptera bonasus* were collected from coastal Mexican waters during May, 2000 to January, 2004: 5 specimens were collected at Ciudad del Carmen, Campeche, Mexico (18° 37' 58" N, 91° 49' 57" W), 1 at Champotón, Campeche (19° 21' N, 90° 54' W), 1 at Isla Contoy, Quintana Roo (20° 48' N, 86° 47' W). The external body surface of each stingray was examined using a magnifying glass and each gill arch was excised, placed in a Petri dish with 0.6% sodium chloride saline solution (Pritchard and Kruse, 1982), and examined using a stereomicroscope. Helminths were removed from gill lamellae and transferred to dishes containing 0.6% sodium...
chloride saline solution. Helminths were lightly flattened under slight coverslip pressure and fixed with Alcohol-Formalin-Acetic Acid at room temperature. Specimens were stained with Gomori’s trichrome (5 worms), Mayer’s carmalum (2 worms), and Delafield’s haematoxylin (4 worms). All specimens were dehydrated in an ethanol series, cleared in methyl salicylate, and mounted in Canada balsam. Specimens were examined using a compound photomicroscope equipped with bright field and Nomarski optics and drawings were made with the aid of a drawing tube. Measurements of curved organs follow the curve. Measurements were made using an ocular micrometer; all measurements are given in micrometers as the mean followed in parentheses by the range and the number of structures measured. Terminology for structures of the haptor follows that of Chisholm et al. (1995) and Chisholm and Whittington (2001). Specimens were deposited in the Colección Nacional de Helmintos (CNHE), Instituto de Biología, Universidad Nacional Autónoma de México, Ciudad de México, Mexico; the University of Nebraska State Museum, Harold W. Manter Laboratory (HWML), Division of Parasitology, Lincoln, Nebraska; and the State Museum, Harold W. Manter Laboratory (HWML), Centre de Investigaciones Biológicas, Universidad Autónoma del Estado de Hidalgo, Pachuca, Hidalgo, Mexico. Three paratypes of *E. occultum* (G 217994-217996) from the Queensland Museum, South Brisbane, Queensland, Australia, were examined.

**Description**

*Euzetia lamothei* n. sp. (Figs. 1-3)

Measurements based on 11 lightly flattened specimens. Total body (excluding haptor) 405 (286-498, n=7) long, by 178 (137-259, n=7) wide at level of anterior part of testis. Haptor oval 210 (178-257, n=7) long, 234 (106-369, n=9) wide with 1 central and 10 peripheral loculi, additional loculus present on either side of central loculus (Figs. 1A and 2A). Unsclerotized ridge on septa of loculi; surface of loculus present on either side of central loculus (Figs. 1A-B). Egg tetrahedral, side length 73 (54-86, n=3) by 51 (38-81, n=10) long, 51 (38-81, n=10) wide, with 6 to 8 transversal muscular bands (Figs. 1A-B). Eye-spots in the form of dispersed pigment granules located anterodorsal to pharynx. Intestinal ceca without diverticula, reaching to posterior portion of body proper, not confluent posteriorly. Cecal bifurcation 130 (101-185, n=5) from anterior end of body. Testis single, spherical to oval, 70 (46-86, n=8) long and 80 (56-114, n=8) wide (Fig. 1B). Vas deferens sigmoid, dilated throughout length, arising from right side of testis, loosely coiled and running anteriorly, dorsal to transverse vitelline ducts, curved toward left side of body, crossing to right side posterior to genital pore. Anterior portion of vas deferens widened to form elongate seminal vesicle that enters ejaculatory bulb (Figs. 1B, 3B). Ejaculatory bulb spherical, 22 (17-27, n=6) long and 18 (14-22, n=7) wide, with 2 prominent spherical chambers (Figs. 1B, 3B). Accessory glands associated with ejaculatory bulb not found. Male copulatory organ (Figs. 1A-B, 3B) sclerotized, in form of small funnel, 20 (16-29, n=4) long and 9 (4-13, n=4) wide at junction of ejaculatory bulb, with stem of funnel, distal portion, doubled back over proximal portion (Fig. 3B); accessory piece absent. Common genital pore located at posterior end of pharynx (Fig. 1B). Ovary elongate, 68 (31-119, n=6) long, V-shaped, encircling right intestinal cecum dorsoventrally and narrowing to form oviduct. Oviduct receives duct from seminal receptacle and common vitelline duct and joins oötype. Mehlis’ gland not prominent, entering posterior portion of oötype. Oötype 61 (31-117, n=5) long and 44 (17-71, n=5) wide, opening medially at unarmed common genital pore. Vaginal pore located 128 (111-142, n=3) from anterior end of body. Vagina muscular, unsclerotized, in the shape of an elongate sac, 41 (28-67, n=7) long and 11 (6-19, n=7) wide at maximum width. Proximal portion of vagina leading to a small, oval vaginal chamber, unsclerotized, 19 (11-36, n=4) long and 9 (5-12, n=4) wide (Figs. 1B, 3C-D). Narrow duct exiting posterior portion of vaginal chamber, connected to spherical seminal receptacle. Receptacle 28 (15-45, n=11) long and 26 (18-33, n=11) wide (Fig. 1B). Spermatophores not observed. Vitellaria extending from level of mid-portion of pharynx to posterior portion of body proper. Transverse vitelline ducts united slightly to right of midbody, common vitelline duct not observed (Fig. 1B). Egg tetrahedral, side length 73 (54-86, n=3) by 53 (50-56, n=3) (measured from egg within oötype); short appendage present.

**Taxonomic summary**

*Type-host:* *Rhinoptera bonasus* (Mitchill, 1815) (Rhinopteridae).

*Type-locality:* Ciudad del Carmen, Campeche, Mexico (18º 37’ 58” N, 91º 49’ 57” W).
Figure 1. Euzetia lamothei n. sp. A, holotype, ventral view. B, detailed ventral view of male and female reproductive system (holotype). Scale bars: A-B = 100 μm.

Site of infection: gills.
Prevalence and intensity: 1 of 1 R. bonasus from the type locality, infected with 11 monogeneans, and 1 of 7 (14%) from the Yucatan Peninsula.
Type specimens: holotype CHNE-6067; paratypes CHNE-6068, HWML-48817, 48818, CHE-P-00056.
Etymology: the specific epithet honors Dr. Rafael Lamothe-Argumedo, Instituto de Biología, Universidad Nacional Autónoma de México, Ciudad de México, Mexico, for his lifelong work and advances in the knowledge of the Monogenea.

Remarks

The subfamily Euzetini was erected by Chisholm and Whittington (2001) for Euzetia, which to date is the only genus assigned to the subfamily. The haptor of Euzetia is similar to that of Decacotylinae in having 1 central and 10 peripheral loculi. However, Euzetia can be distinguished from all other Monocotylidae by a unique feature of the haptor, which has an additional loculus on either side of the central loculus. Specimens of E. lamothei n. sp., like those of E. occultum, have 1 central loculus with 2 flanking loculi and 10 peripheral loculi.

Generally the new species is much smaller than E. occultum. The body of E. lamothei is smaller than that of E. occultum (405 long, 178 wide vs. 1029 long, 674 wide, respectively), as is the haptor (210 long, 234 wide vs. 613 long, 820 wide), pharynx (66 long, 51 wide vs. 152 long, 115 wide), ejaculatory bulb (22 long vs. 34 long), male copulatory organ (20 long vs. 22 long), seminal receptacle (28 long vs. 46 long), and the egg (73 long vs.
Figure 2. *Euzetia lamothei* n. sp. A, haptor, ventral view. B, hamuli (note: not in profile). C, hooklets. Scale bars: A = 100 μm; B-C = 10 μm.

Figure 3. *Euzetia lamothei* n. sp. A, anterior end, ventral view, openings of ducts of anterior glands and papillae surrounding the openings. B, male copulatory complex showing male copulatory organ (mco), ejaculatory bulb (eb) with spherical internal chambers (ic), and seminal vesicle (sv). C, vagina with vaginal pore (vp) and vaginal chamber (vc) (paratype CNHE-6068). D, vagina with vaginal pore (vp) and vaginal chamber (vc) (note: vagina and vaginal chamber contracted) (paratype CHE-P-00056). Scale bars: A-D = 10 μm.
83 long). Finally, *E. lamothei* can be distinguished from *E. occultum*, the only other species currently assigned to the genus, by having the male copulatory organ in the form of a bent cone, the presence of a large sigmoid-shaped vas deferens, and the presence of a vaginal chamber. The copulatory organ of *E. occultum* is a shorter, sclerotized tube that is not bent, the seminal vesicle is small, the vas deferens is an elongated coil having many turns, and the vaginal chamber is absent.

**Discussion**

At present, only 2 species of *Euzetia* have been described. In addition to the species described by Chisholm and Whittington (2001) (*E. occultum* from *Rhinoptera neglecta* Ogiby, 1912 in Australia), these authors based on an unpublished observation made by L. Euzet, suggested that *Euzetia* may include at least 4 species considering 3 undescribed species: 1 from *Rhinoptera steindachneri* Evermann and Jenkins, 1891 from the Pacific Coast of Baja California Sur, Mexico, 1 from *Rhinoptera marginata* (Geoffroy-Saint-Hilaire, 1817) (Geoffroy-Saint-Hilaire, 1817) from Senegal, and 1 from *R. bonasus* from Senegal. Three individuals of *R. steindachneri* from Mazatlán, Sinaloa (23° 12' 17.9" N; 106° 24’ 36.8” W), located at approximately the same latitude and slightly east of the southern tip of Baja California, were examined using the methods described above but no specimens of *Euzetia* were found.

There are only 2 published records of monocotylids from *Rhinoptera*, although members of that genus have been examined previously during various studies with the intent of recovering monogeneans, including but not limited to those of Hargis (1955) and Chisholm and Whittington (2001). The only other monogenean that has been reported from *R. bonasus* is *Benedenella posterocolpa* (Hargis, 1955) Yamaguti 1963 (see Pulido-Flores and Mons, 2005). Eight species of Monocotylidae have been reported from Mexico: *Anoplocotyloides papillatus* (Doran, 1953) Young, 1967 and *Spinitrus mexicana* Bravo-Hollis, 1969 from *Rhinobatos glaucostigma* Jordan and Gilbert, 1883 (Mazatlán, Sinaloa) (Bravo-Hollis, 1969); *S. zapiterygis* Gómez del Prado and Euzet, 1999 from *Zapiteryx exasperata* Jordan and Gilbert, 1880 (Puerto Viejo and Bahía Almejas, Baja California Sur) (Gómez del Prado and Euzet, 1999); *Dendromonocotyle cortesi* Bravo-Hollis, 1969 from mantarraya gris (Isla Rasa and Golfo de California) (Bravo-Hollis, 1969); *Decacotyle floridana* (Pratt, 1910) Chisholm and Whittington, 1998 (= *Heterocotyle acutobatis* Hargis, 1955) from *Aetobatis narinari* Euphasen, 1790 (Ciudad del Carmen, Campeche, and Holbox, Quintana Roo) (Lamothe et al., 1997; Pulido-Flores and Mons, 2005); *Calicotyle californiensis* Bullard and Overstreet, 2000 from *Mustelus californicus* Gill, 1894 (Bahía de los Ángeles, Golfo de California) (Bullard and Overstreet, 2000); *C. urobatis* Bullard and Overstreet, 2000 from *Urobatis halleri* Cooper, 1863 and *U. maculatus* Garman, 1913 (Santa Rosalía, Puertoecitos, San Franciscoquito, and Bahía de los Ángeles, Golfo de California) (Bullard and Overstreet, 2000); and *Dendromonocotyle octodiscus* Hargis, 1955 from *Dasyatis americana* Hildebrand and Schroeder, 1928 (Blanquizal, Quintana Roo) and *U. jamaicensis* (Cuvier, 1816) McEachran and Fechhelm, 1998) (Xcalak, Isla Contoy, El Paso de los Cedros, Cozumel, Quintana Roo, and Río Lagartos, Yucatán (Pulido-Flores and Mons, 2005). Pulido-Flores (2001) provided a complete list of the monogeneans reported up to that time from Mexican stingrays. *Euzetia lamothei* n. sp. is the first species of *Euzetia* and the ninth species of Monocotylidae to be reported from Mexico.

In their revision of Calicotylinae Monticelli, 1903, Chisholm et al. (1997) defined members of the subfamily as being distinct in having an ejaculatory bulb with 2 spherical internal chambers, a condition identified as apomorphic by Chisholm et al. (1995). Since those works, U-shaped internal chambers have been described in the ejaculatory bulb of *H. mokhtarae* Neifar, Euzet and Ben Hassine, 1999, (Heterocotylinae Chisholm, Wheeler and Beverly-Burton, 1995: see Neifar et al., 1999). The 2 species of *Euzetia* that have been described have an ejaculatory bulb with 2 spherical internal chambers (1 pair of chambers) (Fig. 1B, 3B). It is unclear if this character could alter the phylogenetic hypothesis proposed by Chisholm el al. (1995) for the Monocotylidae.

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